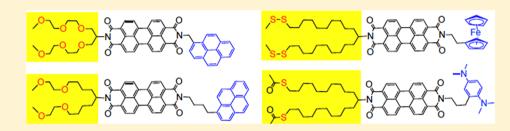
Synthesis of Donor- σ -Perylenebisimide-Acceptor Molecules Having PEG Swallowtails and Sulfur Anchors

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Supporting Information



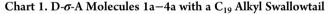
ABSTRACT: Donor- σ -Acceptor (D- σ -A) molecules, arrayed in a monolayer between electrodes, can serve as molecular rectifiers. Using perylene-3,4,9,10-tetracarboxylic bisimide (PBI) as the acceptor allows the attachment of the donor group to one imide nitrogen and a solubilizing swallowtail, normally a long (e.g., C₁₉) alkane connected at midchain, on the other. Such an alkyl tail facilitates the formation of Langmuir–Blodgett (LB) monolayers. We have employed several modified swallowtails to make new D- σ -A molecules: poly(ethylene glycol) (PEG) swallowtails with 6 ether oxygens or with 4 ether oxygens to promote hydrophilicity in orienting LB monolayers, and alkyl swallowtails ending with sulfur anchors (thioacetate, thiol, or methyl disulfide) to stabilize attachment of the D- σ -A molecules to gold electrodes. The preparation and characterization of D- σ -A molecules containing combinations of these swallowtails with pyrene, ferrocene, and tetramethylphenylenediamine donor groups is described.

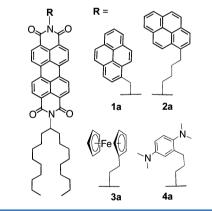
INTRODUCTION

Rectification (the unidirectional flow of current) through a Donor- σ -Acceptor (D- σ -A) molecule, where D is an electrondonating group, A is an electron-accepting group, and σ is a separating bridge of several bonds,^{1–3} was first proposed by Aviram and Ratner in 1974.⁴ Such organic molecular rectifiers have attracted attention as possible components of nanocircuits that could replace present-day silicon-based semiconductor devices.⁵

In the past quarter century, both experimental⁶⁻⁸ and theoretical^{9,10} studies have described rectifying behavior of D- σ -A molecules. To measure current through a potential D- σ -A rectifier, it is convenient to assemble a monolayer of the molecules and interrogate a collection of the monolayer molecules aligned in parallel. Monolayers may be formed by Langmuir–Blodgett (LB)^{6,11–13} or self-assembled monolayer (SAM)^{14–18} techniques. LB monolayers form when molecules organize themselves on the surface of an aqueous/air interface. To obtain a well-ordered LB monolayer, suitable for lifting onto a bottom electrode for electrical studies, an alkyl tail typically is attached to one end of the D- σ -A molecule to give it a hydrophobic end that will prefer to be in the air.¹ In contrast, formation of a SAM typically involves attaching a sulfur anchor to one end of the D- σ -A molecule, so that it will coordinate to the bottom Au electrode. The top electrode can be a scanning probe or can be created by evaporative metal techniques.⁵

We have previously described the synthesis $(1a-4a, Chart 1)^1$ and LB film formation and monolayer electrical properties





 $(1a-3a)^6$ of several D- σ -A molecules based on perylene-3,4,9,10-tetracarboxylic bisimide (PBI) as the acceptor. In order to provide adequate solubility and the required hydrophobic alkyl tail for LB film formation, a "swallowtail", a long alkyl chain connected at its center,^{1,19} is attached to one imide end. (The letter **a** designates the C₁₉ alkyl swallowtail shown; other letters will designate different swallowtail targets.) The other imide end holds the σ bridge to the donor group: pyrene (1a, 2a), ferrocene (3a), or tetramethylphenylenediamine (4a). A

Received: August 13, 2012 Published: October 10, 2012 well-packed LB film is not formed from 2a, but LB monolayers of 1a and 3a, arranged between two electrodes in a "AulD- σ -Al Au" sandwich, rectify electric current; 3a has a rectification ratio between 14 and 28 at ± 1 V.⁶

In 1a-4a, the donor end can be considered more polar than the C_{19} alkyl swallowtail end (even in the case of the donor pyrene, which is not particularly polar). It is therefore assumed that the donor end will prefer the aqueous subphase of the LB trough and will comprise the bottom of the monolayer, while the swallowtail end will be in air and comprise the top face (Figure 1a).¹ To have control of the direction of electron

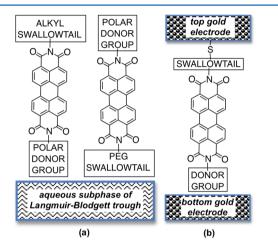


Figure 1. (a) Proposed orientation of swallowtailed D- σ -A molecules on an LB trough. (b) Stabilization with sulfur anchors of swallowtailed D- σ -A molecules in one of the two possible orientations.

transport across the monolayer, we wanted to be able to reverse the orientation of the LB film by changing the polarity of the swallowtail. Swallowtails with hydrophilic poly(ethylene glycol) (PEG) arms instead of alkyl arms would be more hydrophilic than the donors and might thereby reverse the film's orientation and its direction of rectification (Figure 1a). Fullerenes decorated with a PEG hemisphere^{20a} and diblock copolymers with a PEG block^{20b} can be oriented into Langmuir films in this way. In addition, PEG ether oxygens would provide a less-insulating layer compared to the hydrocarbon swallowtails, and this might allow for a greater tunneling current through the monolayer.

PEG tails are widely used to improve solubility,²¹ and like alkyl swallowtails, various PEG tails have been used to improve the solubility of perylenebisimides.²² PEG tails can impart liquid crystalline characteristics to PBI compounds,²³ which improve $\pi - \pi$ stacking, allowing dynamic reorganization and easier processing. Cormier and Gregg²³ reported liquidcrystalline PBIs with branched propylimide-oligo(oxyethylene) or phenethylimide-oligo(oxyethylene) side chains, while Murray et al.²⁴ synthesized PBIs with short poly(ethylene oxide) tails. We planned to use a PEG swallowtail corresponding to the skeleton of the C₁₉ alkyl swallowtail.

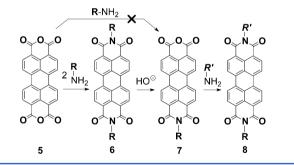
We are also interested in stabilizing the monolayers after their assembly into AulD- σ -AlAu sandwiches. For example, the rectification ratios of **1a** and **3a** decrease on repeated voltage cycling, a phenomenon that is frequently seen in such systems.⁵ A possible explanation for this attenuation is that molecules within the monolayer may flip their orientation in response to the cycling electric field. The incorporation of sulfur anchor groups at the ends of the swallowtail arms would inhibit the putative molecular flipping by coordinating the sulfur anchors to the gold electrode (Figure 1b). Thiol anchors have a strong, specific affinity toward gold^{17,18} and form good SAMs, but the -SH group tends to be unstable, so thiols are often protected as thioesters (e.g., S-acetyl^{25a}) or cyanoethyl thioethers^{25b} and deprotected just before use. This strategy has been used to stabilize rectifying layers of a D- π -A molecule with an SAc anchor at the end of single alkyl tails of different lengths.^{26a,b} S-Acetyl anchors themselves are capable of generating SAMs on gold, but the assembly process is slow and the properties of the resulting films differ from those of thiol-derived SAMs.^{26c} Highquality SAMs can be formed from disulfides, but if these are unsymmetrical (e.g., RS-SCH₃), the coverage will differ from SAMs made from the corresponding thiols (RSH) or thioacetates (RSAc). Since SAMs to gold can be formed from all three of the above-mentioned sulfur groups, they are potential anchors for our D- σ -A molecules.

Here we report the synthesis of several D- σ -A molecules with PEG swallowtails, intended to reverse the orientation of the monolayers respective to the monolayers of molecules 1a-4aand thereby reverse the direction of rectification, and D- σ -A molecules with sulfur anchors at the ends of alkyl swallowtails, to stabilize AuID- σ -AlAu assemblies and sustain their rectification ratios, as well as making D- σ -A molecules capable of SAM formation. The formation and properties of monolayers made from these molecules and the investigation of their electrical properties will be described elsewhere.

RESULTS AND DISCUSSION

Unsymmetrical PBIs (8) are prepared from commercially available perylenetetracarboxylic bisanhydride 5 (Scheme 1). In

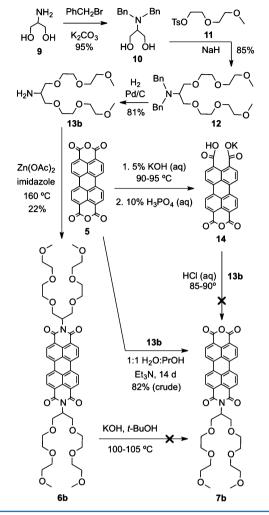
Scheme 1. General Procedure To Make Unsymmetrical PBIs 8



principle, one anhydride group of **5** could condense with a primary amine, R-NH₂, to make the monoimide monoanhydride 7, followed by condensation of the other anhydride group with a second amine, R'-NH₂, to make **8**. In practice, the first condensation under typical conditions (e.g., molten imidazole and Zn(OAc)₂) usually affords only symmetrical bisimide **6**,^{1,27,28} which is nicely soluble if R-NH₂ is a swallowtail. This bisimide is then semihydrolyzed to the imide anhydride 7 before introduction of the second amine,^{1,29} which in our system adds the donor group. The preparation of appropriate swallowtail amines, R-NH₂, was therefore the first step in making the intermediates 7 required for the synthesis of the D- σ -A targets.

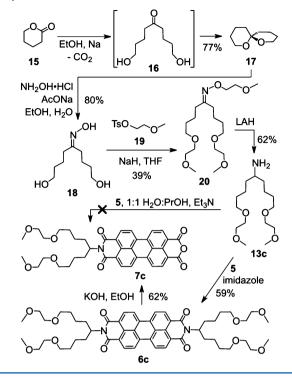
Swallowtail Amines (13). The PEG swallowtail amine 13b (Scheme 2) was prepared by double Williamson alkylation of N,N-dibenzyl-protected serinol 10 with diethylene glycol monomethyl ether tosylate 11, followed by hydrogenolysis of

Scheme 2. Synthesis of PEG Swallowtail Imide Anhydride 7b



the benzyl groups of the protected PEG amine 12, as described previously.²¹ We had converted 13b to the symmetrical, watersoluble PBI 6b as shown in Scheme 2; the water solubility of 6b supported the idea that a single PEG swallowtail in a PBI would be hydrophilic enough to seek the water side of an LB monolayer. However, the troublesome solubility profile of the crucial imide anhydride 7b prepared from 13b (see below) prompted us to attempt an alternative, a "hybrid" swallowtail, 13c, that ended in a PEG segment for hydrophilicity in an LB film but had alkyl character near to the imide nitrogen to promote better solubility in organic solvents (Scheme 3). We anticipated making amine 13c from dihydroxyketone 16, the product of a Claisen condensation of δ -valerolactone 15. This condensation, however, is known to give only the spiroketal 17 (a pheromone of the female olive fly).³⁰ Ketal 17 is a thermodynamic sink for 16, and various attempts to make 16 invariably produce 17 instead.³¹⁻³⁶ No reactions have been reported for 17; this is attributed to its stable nature. We adopted Baker's procedure³⁰ for synthesizing 17 using a onestep Claisen condensation of 15, and we then succeeded in converting 17 to its oxime 18 in good yield, essentially capturing 17 as it equilibrated with an open-chain form. Williamson alkylation of the hydroxyls of 18 with methoxyethyl tosylate 19 was accompanied by oxime alkylation, giving the triple-alkylated 20. Alkylation of the oxime hydroxyl was not a

Scheme 3. Synthesis of Mixed-PEG Swallowtail Imide Anhydride 7c

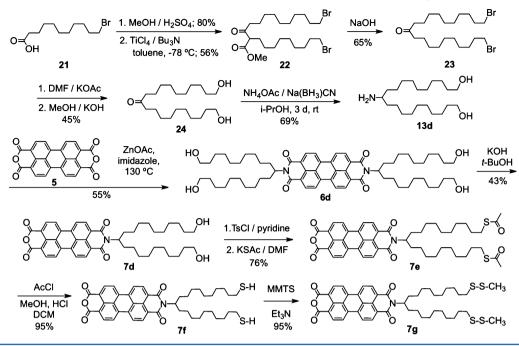


concern, however, since LiAlH_4^{37} in refluxing THF reduced the *N*-alkoxy group to the desired primary amine **13c**.

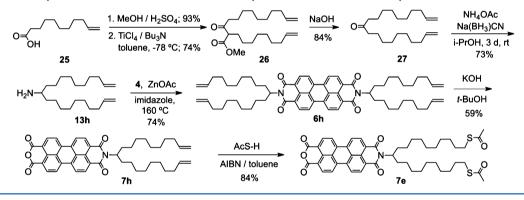
Our first attempt to obtain alkyl swallowtails with sulfur anchors was *via* transformation of terminal hydroxyl groups (Scheme 4). 11-Bromoundecanoic acid (**21**) was esterified and then underwent TiCl₄-catalyzed Claisen condensation³⁸ and decarboxylation to make the known dibromoketone **23**. Attempts to make the oxime of **23** were unsuccessful, presumably because of side reaction at the bromines, so the bromines were converted to hydroxyls by acetate substitution and methanolysis, yielding the dihydroxyketone **24**. In contrast to **17**, the intramolecular ketal is not favored in this larger system. Reductive amination of **24** gave the desired amine **13d**. This amine was converted to its imide anhydride **7d** *via* the bisimide **6d** before introduction of the thioacetate group (see below).

We also pursued an alternative route to obtain alkyl swallowtails with sulfur anchors that involved a swallowtail amine with terminal vinyl groups, 13h (Scheme 5). 11-Undecenoic acid (25) underwent esterification, followed by Claisen condensation as before to give the divinyl ketone 27^{38} and then reductive amination to give 13h. Again, introduction of the sulfur groups was delayed (see below) until after formation of the corresponding imide anhydride 7h, prepared *via* the bisimide 6h.

Perylenetetracarboxylic Monoimide Monoanhydrides (7). The general approach for making imide anhydrides outlined in Scheme 1 worked well for amines 13c, 13d, and 13h. They condensed with bisanhydride 5 to make symmetrical bisimides 6c, 6d, and 6h, followed by semihydrolysis^{1,29,40} to the monoimide monoanhydrides 7c, 7d, and 7h, as shown in Schemes 3, 4, and 5, respectively. In the case of 13d, the standard conditions for making the bisimide were too harsh, but lowering the reaction temperature from 160° to 130° resulted in a satisfactory yield of 6d. Scheme 4. Synthesis of Sulfur-Anchored, Alkyl-Swallowtailed Imide Anhydrides 7e, 7f, and 7g



Scheme 5. Alternate Synthesis of Thioacetate-Anchored, Alkyl-Swallowtailed Imide Anhydride 7e



The swallowtails of **6d** and **6h** closely resemble alkyl swallowtails, differing only in the presence of hydroxyl (**6d**) or vinyl (**6h**) groups at the distal ends of the swallowtail arms. A modification of the procedure was needed to prepare intermediate **7c** (Scheme 3), in which the swallowtail is PEGlike for the five distal skeletal atoms of each arm, but alkyl for the four skeletal atoms of each arm closest to the perylenebisimide. The standard semihydrolysis conditions of KOH in *t*-BuOH were unsuccessful with **6c**, because the reaction was too fast and both imides were hydrolyzed. A change of solvent to ethanol, however, gave a satisfactory yield (62%) of **7c**. As hoped for, monoimide monoanhydride **7c**, in contrast to PEG analogue **7b**, was soluble in organic solvents, and hence we could easily verify its structure using NMR.

Difficulties arose when we attempted to prepare the monoimide monoanhydride containing a full PEG swallowtail (7b) by the standard method. The PEG-amine 13b converted easily to PBI 6b (Scheme 2). However, in contrast to the alkyl-like monoimide monoanhydrides 7c, 7d, and 7h, which had been soluble in organic solvents and therefore easily isolated following semihydrolysis, 7b was insoluble in common organic solvents, complicating its purification. We were unable to

isolate adequate amounts of 7b by this method and therefore turned to alternate methods.

In our hands, Tröster's method⁴¹ of making imide anhydrides directly from the monoanhydride monopotassium salt (14) of 5 was not successful with the PEG amine 13b (Scheme 2). The crude reaction mixture failed to show anhydride carbonyl peaks in the infrared spectrum. (We also tried Tröster's salt 14 with the dihydroxyalkyl swallowtail amine 13d but obtained an intractable polar product.) Pasaogullari and Demuth's modified procedure⁴² employing Tröster's salt⁴¹ also failed in preparing 7b.

Nagao and Misono's method (lengthy refluxing of **5** with a primary amine in a $H_2O/PrOH/Et_3N$ mixture⁴³) was able to convert PEG amine **13b** into imide anhydride **7b** (Scheme 2). However, compound **7b** was not soluble in common organic solvents, although it was soluble in basic solvents such as morpholine, pyridine, and diethylamine. Due to its poor solubility, it was difficult to verify its purity following purification attempts by column chromatography; consistent with the thermal instability of PEGs,⁴⁴ sublimation was also unsuccessful. Crude **7b** was therefore used to carry the synthesis forward. The Nagao–Misono method is generally less useful for large alkyl swallowtails, presumably because of

their poor solubility in the polar solvents, but we thought the half-PEG, water-soluble amine **13c** might be a worthy substrate. However, only trace amounts of monoimide monoanhydride **7c** were obtained with this method (Scheme 3).

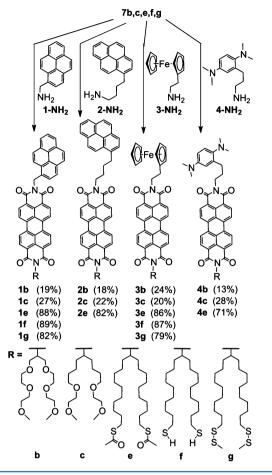
Introduction of Sulfur End Groups into Swallowtailed Monoimide Monoanhydrides. With the hydroxyl-tipped alkyl-swallowtailed imide anhydride 7d, tosylation and displacement with thioacetate afforded the thioacetate-tipped 7e (Scheme 4). This compound could also be obtained from the vinyl-tipped imide anhydride 7h by a double radical-chain addition of thioacetic acid⁴⁵ to the terminal double bonds (Scheme 5), a reaction that required dry solvents and an inert atmosphere to obtain good yields. The route to thioacetyltipped 7e was shorter *via* the divinyl swallowtail 13h compared to the diol 13d, and the overall yield from starting C₁₁ acid was much better for the divinyl route (21%) compared to the diol route (2%).

The bisthioacetate imide anhydride 7e was converted to the dithiol imide anhydride 7f by treatment with acetyl chloride in MeOH,⁴⁶ which provided an acidic environment that promoted methanolysis (Scheme 4). In turn, the dithiol 7f was converted to the bis(methyldisulfide) 7g by treatment with methyl methanethiosulfonate.⁴⁷

Conversion of Monoimide Monoanhydrides 7 to Unsymmetrical Bisimides. The condensation of pervlene monoimide monoanhydrides 7 with donor amines pyrenylmethyl $(1-NH_2)$, pyrenylbutyl $(2-NH_2)$, ferrocenylethyl (3- NH_2), or TMPDA-propyl (4- NH_2) to make unsymmetrical bisimides was straightforward (Scheme 6). Target D- σ -A molecules 1b, 2b, 3b, and 4b, with the full PEG swallowtail, were made by refluxing crude PEG imide anhydride 7b and the appropriate donor amine in pyridine overnight. In contrast to the precursor imide anhydride 7b, all of these PBI targets were soluble in chloroform, and recrystallization (CHCl₃/hexane) followed by column chromatography (99:1 CHCl₃/MeOH) resulted in pure products. Target D- σ -A molecules 1c, 2c, 3c, and 4c with mixed alkyl-PEG swallowtails were made by refluxing crude imide anhydride 7c with the appropriate donor amine in toluene. Target D- σ -A molecules 1e, 2e, 3e, and 4e, containing thioacetate-anchored alkyl swallowtails, were made by refluxing crude imide anhydride 7e with the appropriate donor amine in toluene; in the case of the reaction of $4-NH_2$, slow addition of the amine was required to suppress side reaction of the thioacetates. We decided to utilize the other two, less widely used sulfur anchor groups, thiol f and disulfide g, with two of the donors, 1-NH₂ and 3-NH₂. Condensation of imide anhydrides 7f and 7g with these amines gave targets 1f, 1g, 3f, and 3g.

Attempted Direct Condensation to Unsymmetrical Bisimides. Although the final condensations shown in Scheme 6 were straightforward, the yields were disappointing (around 20%) for the PEG and mixed-PEG swallowtails **b** and **c**, compared to the alkyl swallowtails **e**, **f**, and **g** (around 80%). We therefore attempted an alternative "direct" condensation by treating the bisanhydride **5** with PEG swallowtail amine **13b** combined with either **1-NH**₂ or **3-NH**₂. Such attempts to avoid the isolation of the monoimide monoanhydride intermediate are usually not very successful:³² even if both amines have similar reactivity, there will be two symmetrical bisimide products along with the desired crossed product, and these will be difficult to separate. However, if only one of the amines is a swallowtail may impart unusual solubility to bisimides

Scheme 6. Formation of Unsymmetrical PBI D-σ-A Targets



containing it;¹⁴ in this case, the direct procedure is much simpler and shorter than the conventional route of Scheme 1. In the event, treatment of **5** with 5- to 10-fold excesses of both donor amine and **13b** in either hot pyridine or imidazole for 1.5 d resulted in crude products from which no color was extractable into CHCl₃. Since the target PBIs are deep red and soluble, the procedure was unsuccessful.

Carbonyl ¹³C Peaks. For PBIs with alkyl swallowtails (e.g., 1a-4a), slow rotation about the N-CH bond, along with a preferred conformation that places the CH bond in the plane of the carbonyls, makes the two carbonyls nonequivalent on the NMR time scale. This gives rise to two weak carbonyl peaks in the ¹³C NMR spectrum.¹ We observed similar nonequivalence in the symmetrical PBIs that had alkyl tails tipped with hydroxyl (6d) and vinyl (6h) groups and in imide anhydrides containing those swallowtails (7d, 7h) or alkyl swallowtails tipped with sulfur-containing groups (7e, 7f, 7g). Changing to a PEG swallowtail apparently increases the rotation rate about the N-CH bond, since only one signal was observed for the swallowtail carbonyls in the bisimides $6b^{21}$ and 1b-4b. In agreement, MM2 calculations of a model swallowtail with oxygens γ to the N predicted a rotation barrier of 8.5 kcal/mol, lower than the prediction for an alkyl swallowtail of 11.8 kcal/ mol.¹ The hybrid alkyl-PEG swallowtail would be expected to be dominated by its alkyl portion, and the symmetrical bisimide 6c indeed showed two carbonyl peaks. The results were not so clear-cut for the imide anhydride 7c and the unsymmetrical bisimides 1c-4c. One weak carbonyl peak could be one of a pair, with the second peak coincident with a larger carbonyl

peak from the other end of the molecule. Interestingly, this same situation was observed in unsymmetrical bisimides with sulfur-tipped swallowtails (1e-4e, 1g, 3g), in which the nature of the swallowtail is clearly alkyl.

CONCLUSIONS

We report here the synthesis of 16 new D- σ -A molecules, combining the PBI acceptor with three different donor groups (with two different tether lengths for the pyrene donor) and five different swallowtails. We tried several approaches for constructing the required unsymmetrical perylene bisimide architecture, and it is interesting that the approaches gave such varied results. The Tröster salt and direct condensation approaches, useful in some systems, were completely unsuccessful here. The familiar semihydrolysis of PBIs 6 to imide anhydrides 7 shown in Scheme 1 was the preferred route for alkyl-like swallowtails, including the mixed alkyl-PEG 13c. In contrast, the full PEG swallowtail amine 13b was best managed with the basic equilibration method of Nagao and Misono.

EXPERIMENTAL SECTION

General Methods. Oxygen- and/or moisture-sensitive reactions were carried out under N_2 in glassware that was dried under vacuum and purged with N_2 prior to use. Reagents and solvents were purchased from commercial vendors and used as received. ¹H and ¹³C NMR spectra were recorded on 300 or 500 MHz spectrometers at 25 °C. IR spectra were obtained with an FTIR spectrometer. Chromatography was performed using 60 Å silica gel.

2-(N,N-Dibenzylamino)-1,3-propanediol (10). 2-Aminopropane-1,3-diol (5.21 g, 57.2 mmol) and anhydr potassium carbonate (24.5 g, 177 mmol) were suspended in ethanol (200 mL). Benzyl bromide (30.3 g, 177 mmol) was added dropwise over 30 min with stirring. After the mixture had refluxed for 3 h, solids were separated by filtration, and volatiles were removed under reduced pressure. The residue was dissolved in EtOAc; the solution was washed with water, aq NaHCO₃, and brine and dried with anhyd MgSO₄; and the solvent was removed under reduced pressure. The crude mixture was purified by recrystallization from 1:1.2 benzene/hexane to give 14.7 g (95%) of white needles. ¹H NMR (300 MHz, CDCl₃) δ 7.40–7.25 (m, 10 H), 3.74 (s, 4 H), 3.69 (m, 2 H), 3.59 (m, 2 H), 2.99 (m, 3 H). ¹³C NMR (75 MHz, CDCl₃) δ 139.2, 128.8, 128.3, 127.3, 59.8, 59.7, 53.9. NMR spectra are in agreement with the literature.^{21,48}

2-(2-Methoxyethoxy) Ethyl Tosylate (11). To a solution of diethyleneglycol monomethyl ether (4.35 g, 36.2 mmol) in 15 mL of THF, cooled at 0 °C, was added NaOH (2.46 g, 61.5 mmol) dissolved in 15 mL of water with vigorous stirring. To this mixture was added dropwise a solution of tosyl chloride (8.28 g, 43.4 mmol) in 15 mL of THF over 10 min at 0 °C. The reaction mixture was then raised to rt and stirred for 1 h under N₂. It was then extracted twice with 50 mL of diethyl ether, and the organic layer was washed with 1 M aq NaOH and water and dried with anhyd MgSO₄. Solvent was removed under reduced pressure to yield 9.0 g (91%) of colorless liquid 11. ¹H NMR (300 MHz, CDCl₃) δ 7.81 (d, *J* = 6.0 Hz, 2H), 7.31 (d, *J* = 6.0 Hz, 2H), 3.47 (m, 2H), 3.34 (s, 3H), 2.42 (s, 3H). NMR spectrum is in agreement with the literature.⁴⁹

N,N-Dibenzyl-2,5,8,12,15,18-hexaoxa-10-nonadecanamine (12). THF (50.0 mL) was added to NaH (60% suspension in mineral oil) (0.80 g, 33.4 mmol), and the mixture was stirred to make a suspension. A solution of 10 (4.12 g, 15.1 mmol) in 15 mL of THF was then added dropwise under nitrogen, and after 30 min of stirring, a solution of 11 (9.16 g, 33.4 mmol) in 20 mL of THF was added dropwise. The reaction mixture was refluxed overnight, cooled, and treated carefully with MeOH to destroy excess NaH. The solvent was removed under reduced pressure and the resulting brown oil was extracted 4 times with 20 mL portions of hot ether. The combined

ether fractions were concentrated by rotary evaporation to give 5.12 g (71%) of a yellowish liquid, R_f (EtOAc) 0.47. ¹H NMR (300 MHz, CDCl₃) δ 7.39 (m, 4H), 7.28 (m, 4H), 7.20 (m, 2H), 3.78 (s, 4H), 3.55 (m, 20H), 3.37 (s, 6H), 3.10 (m, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 140.8, 128.7, 128.1, 126.7, 72.0, 70.7, 70.6, 70.5, 70.4, 59.0, 56.2, 55.2. NMR spectra are in agreement with the literature.²¹

2,5,8,12,15,18-Hexaoxa-10-nonadecanamine (13b). The *N,N*-dibenzyl PEG-amine **12** (4.00 g, 8.41 mmol) was dissolved in 110 mL of MeOH, and 2.5 g of 10% Pd/C catalyst was added. The mixture was hydrogenated overnight in a Parr shaker at 55 psi of H₂. The reaction mixture was then filtered, and the solvent was removed under reduced pressure to give 2.0 g (81%) of an oil, R_f (72:18:10 CH₂Cl₂/NH₂OH(aq)/MeOH) 0.51. ¹H NMR (300 MHz, CDCl₃) δ 3.60 (m, 12H), 3.51 (m, 6H), 3.35 (m, 8H), 3.17 (m, 1H), 2.04 (s, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 72.0, 71.9, 70.6, 70.56, 70.51, 70.4, 59.1. IR (thin film) 3522, 3364, 3290, 2891 cm ⁻¹. NMR and IR are in agreement with the literature.²¹

N,*N*[']-Di[10-(2,5,8,12,15,18-hexaoxanonadecyl)]perylene-3,4,9,10-bis(dicarboximide) (6b). PEG amine 13b (212 mg, 0.71 mmol), dianhydride 5 (128 mg, 0.32 mmol), imidazole (2.62 g), and Zn(OAc)₂ (catalytic amount) were heated at 165 °C overnight. The reaction mixture solidified upon cooling and was dissolved in MeOH, then run through a Sephadex column to obtain 75 mg (22%) of deepred waxy solid, *R_f* (2:1 MeOH/EtOAc) 0.70. ¹H NMR (300 MHz, CDCl₃) δ 8.63 (m, 4H), 8.58 (m, 4H), 5.72 (m, 2H), 4.19 (m, 4H), 3.99 (m, 4H), 3.72 (m, 4H), 3.61 (m, 12H), 3.56 (m, 8H), 3.42 (m, 8H), 3.28 (s, 12H). ¹³C NMR (75 MHz, CDCl₃) δ 164.0, 134.6, 131.7, 129.7, 126.4, 123.6, 123.2, 72.0, 70.63, 70.62, 70.5, 69.4, 59.1, 52.2. IR (KBr) 1700 (C=O), 1654 (C=O) cm⁻¹. NMR and IR are in agreement with the literature.²¹

N-[10-(2,5,8,12,15,18-Hexaoxanonadecyl)]-3,4,9,10-perylenetetracarboxylic Acid 3,4-Anhydride 9,10-Imide (7b). To a mixture of perylenedianhydride 5 (498 mg, 1.27 mmol) and the PEG swallowtail amine 13b (372 mg, 1.25 mmol) in 40 mL of propanol and water (1:1) was added triethylamine (1.68 g, 16.6 mmol), and the reaction was heated for 15 d at 120 °C. The reaction mixture was treated with excess 5% HCl at 120 °C for 10 min, cooled to room temperature, filtered, and dried to give 0.70 g (82%) of the perylene imide anhydride 7b as a waxy solid. Due to limited solubility, crude material was carried further (see below), and no NMR data was collected. IR (KBr) 1768 (s) (anhydride C=O), 1733 (m) (anhydride C=O), 1693 (s) (imide C=O), 1648 (s) (imide C=O) cm⁻¹. HRMS (TOF MS ES⁺) calcd for $C_{37}H_{35}NO_{11}$ 669.2108; found 669.2109.

1,7-Dioxaspiro[**5.5**]**undecane** (**17**). Sodium metal (1.15 g, 50.0 mmol) was dissolved in 50 mL of ethanol. To this was added dropwise a solution of δ -valerolactone (**15**, 10.0 g, 100 mmol) in 15 mL of ethanol at 0 °C, and then the mixture was refluxed for 5 h, cooled, acidified with 5% HCl, and refluxed again for 30 min. The reaction mixture was then co-distilled with water. The water distillate was extracted with ether, which was then dried over anhyd MgSO₄ and concentrated to give 6.00 g (77%) of clear colorless liquid. ¹H NMR (500 MHz, CDCl₃) δ 3.69 (d × t, *J* = 7.1, 14.0 Hz, 2H), 3.63 (m, 2H), 1.83 (m, 2H), 1.5 (m, 10H), consistent with the literature⁴ values. IR (liquid film) 1652 cm⁻¹.

1,9-Dihydroxy-5-nonanone Oxime (18). 1,7-Dioxaspiro[5.5]undecane (17, 6.00 g, 38.4 mmol), hydroxylamine hydrochloride (2.77 g, 39.9 mmol), and sodium acetate (3.24 g, 39.5 mmol) were added to a mixture of 40 mL of ethanol and 30 mL of water, and the reaction mixture was heated with constant stirring and a closed condenser to prevent the escape of the volatile 17. This reaction was conducted at 40 °C for 6 h, 60 °C for 4 h, and then at 80 °C for 26 h. The reaction mixture was cooled to rt, basified with aq NaHCO₃, and extracted with EtOAc (4 × 20 mL). The extraction was done with warm solvents to encourage the removal of the water-soluble **18**. The organic layer was dried over anhyd MgSO₄ and concentrated to give a viscous, colorless liquid, which was purified by silica gel column chromatography (5:95 MeOH/EtOAc) to give 4.00 g (80%) of a viscous liquid. ¹H NMR (500 MHz, DMSO-*d*₆) δ 10.17 (s, 1H), 4.37 (t, *J* = 5.0 Hz, 2H), 3.40– 3.33 (m, 4H), 2.20 (t, *J* = 8.0 Hz, 2H), 2.11–2.08 (t, *J* = 7.0 Hz, 2H), 1.46–1.39 (m, 8H). ¹³C NMR (125 MHz, DMSO- d_6) δ 159.0, 60.93, 60.91, 33.6, 33.1, 32.6, 27.0, 22.9, 22.2. IR (liquid film) 3400 (s), 1645 (s) cm⁻¹. HRMS-ESI calcd for C₉H₁₉NO₃Na 212.1263; found 212.1255.

N-(2-Methoxyethoxy)-2,5,15,18-tetraoxanonadecan-10-one Oxime (20). 1,9-Dihydroxynonan-5-one oxime (18, 1.50 g, 7.93 mmol) in 20 mL of THF was added dropwise at 0 °C to a solution of NaH (60% in mineral oil, 1.40 g, 35.0 mmol) in 30 mL of THF, and the resulting mixture was stirred for 30 min. To this was added 2methoxyethyl tosylate⁵⁰ (7.80 g, 33.9 mmol) in 20 mL of THF dropwise at 0 °C. Then the reaction mixture was refluxed for 12 h with constant stirring. The reaction mixture was cooled to rt, and MeOH was added dropwise to destroy the excess NaH. The mixture was rotary evaporated, and the resulting crude product was extracted with ether. The organic layer was concentrated to give a yellowish liquid, which then was column chromatographed over silica gel eluting with EtOAc to give 0.70 g (39%) of pure product as an oily liquid. ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3) \delta 4.14 \text{ (t, } J = 4.8 \text{ Hz}, 2\text{H}), 3.62-3.53 \text{ (m, 10H)},$ 3.48 (t, J = 6.6 Hz, 4H), 3.40 (s, 6H), 3.39 (s, 3H), 2.32 (t, J = 7.5 Hz, 2H), 2.17 (t, J = 7.2 Hz, 2H), 1.64–1.51 (m+t, J = 7.8 Hz, 8H). ¹³C NMR (125 MHz, CDCl₃) δ 161.3, 72.5, 72.1, 71.3, 71.27, 71.24, 70.2, 70.1, 59.2, 59.1, 33.9, 29.8, 29.4, 27.9, 23.2, 22.6. IR (liquid film) 1635 (w), 1456 (m) cm $^{-1}$. HRMS (TOF MS ES $^{+})$ calcd for $\rm C_{18}H_{37}NO_6Na$ 386.2519; found 386.2520.

2,5,15,18-Tetraoxanonadecan-10-amine (13c). Compound 18 (700 mg, 1.93 mmol) was dissolved in THF on ice with constant stirring. To this was added LAH (150 mg, 3.95 mmol), and the reaction mixture was stirred for 10 min on ice and 2 d at reflux. The LAH was quenched with EtOAc and then 10% NaOH, and the resulting mixture was stirred for 20 min. The reaction mixture was then filtered and extracted with EtOAc to give a colorless viscous liquid, which was purified by silica gel column chromatography to give 0.35 g (62%) of pure product as an oily liquid. ¹H NMR (300 MHz, CDCl₃) δ 3.60–3.54 (m, 8H), 3.50 (t, *J* = 6.3 Hz, 4H), 3.39 (s, 6H), 3.19 (s, 2H+H₂O) 2.85 (m, 1H), 1.66–1.54 (m, 4H), 1.51–1.41 (m, 8H). ¹³C NMR (125 MHz, CDCl₃) δ 71.9, 71.3, 69.9, 59.0, 51.0, 37.7, 29.6, 22.6. IR (liquid film) 3369 (s), 1592 (m) cm⁻¹. HRMS (TOF MS ES⁺) calcd for C₁₅H₃₃NO₄Na 314.2307; found 314.2283.

N,N'-Di[10-(2,5,15,18-tetraoxanonadecyl)]perylene-3,4,9,10bis(dicarboximide) (6c). A mixture of 13c (4.00 g, 13.7 mmol), 3,4,9,10-perylenetetracarboxylic dianhydride (5, 2.50 g, 6.37 mmol), imidazole (10.0 g), and a catalytic amount of $Zn(OAc)_2$ (127 mg) was heated at 165 °C for 8 h. The reaction mixture solidified upon cooling and was dissolved in CH2Cl2 and washed with 5% HCl (aq) followed by brine solution. The organic layer was dried over anhyd MgSO₄, concentrated, and purified by neutral Al₂O₃ column chromatography (CHCl₃) followed by flash column chromatography over silica gel (1:9 $MeOH/CH_2Cl_2$) to give 3.51 g (59%) of pure product as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.59–8.44 (m, 8H), 5.21–5.16 (m, 2H), 3.51-3.40 (m, 24H), 3.29 (s, 12H), 2.35-2.22 (m, 4H), 1.94-1.84 (m, 4H), 1.65–1.69 (m, 8H), 1.44–1.34 (m, 8H). ¹³C NMR (75 MHz, CDCl₃) δ 164.3, 163.3, 134.2, 131.6, 130.9, 129.3, 126.1, 123.6, 122.9, 122.8, 71.8, 71.1, 69.8, 58.9, 54.3, 32.0, 29.3, 23.4. IR (KBr pellet) 1697 (s), 1655 (s), 1595 (w) cm⁻¹. HRMS (TOF MS ES⁺) calcd for C54H70N2O12Na 961.4827; found 961.4847.

N-[10-(2,5,15,18-Tetraoxanonadecyl)]-3,4,9,10-perylenetetracarboxylic Acid 3,4-Anhydride 9,10-Imide (7c). A solution of 6c (500 mg, 0.532 mmol) in EtOH was treated with KOH as follows, with monitoring by TLC. First 124 mg of KOH (2.21 mmol) was added while the reaction was heated at 55 °C for 1 h, then 240 mg of KOH (4.28 mmol) at 65 °C for 2 h, and finally 250 mg of KOH (4.46 mmol) at 75 °C for 2 h. The reaction mixture was then cooled to rt, acidified with 5% HCl and extracted with CHCl₃. The organic layer was treated with brine, dried over anhyd MgSO₄, and concentrated to give a solid. This was purified by silica gel column chromatography (5:95 MeOH/CHCl₃) to give 220 mg (62%) of pure product as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.69–8.62 (m, 8H), 5.22 (m, 1H), 3.54–3.42 (m, 12H), 3.33 (s, 6H), 2.32–2.28 (m, 2H), 1.91–1.87 (m, 2H), 1.69–1.60 (m, 4H+H₂O), 1.49–1.24 (m, 4H). ¹³C NMR (75 MHz, CDCl₃) δ 163.4, 162.9, 135.3, 134.1, 131.2, 123.3, 123.0, 71.8, 71.2, 69.9, 59.0, 54.5, 32.1, 29.4, 23.5. HRMS (TOF MS ES⁺) calcd for $C_{39}H_{39}NO_9Na$ 688.2523; found 688.2545.

Methyl 13-Bromo-2-(9-bromononyl)-3-oxotridecanoate (22). A solution of 11-bromo undecanoic acid (21) (24.7 g, 93.0 mmol) and concd H_2SO_4 (5 mL) in MeOH (300 mL) was refluxed for 4 h. After most of the solvent was removed under reduced pressure, water was added, and the mixture was extracted with ether. The ether layer was washed with 5% NaHCO₃, water, and brine, dried over anhyd MgSO₄, and concentrated by rotary evaporation to yield 20.9 g (80.0%) of crude methyl undecanoate, ⁵¹ which was used without purification in the next step. ¹H NMR was in agreement with the literature.⁵²

TiCl₄ (8.07 g, 42.5 mmol) in toluene (50 mL) was added to a stirred solution of methyl undecanoate (7.92 g, 28.4 mmol) and Bu₃N (9.46 g, 51.1 mmol) in toluene at -78 °C. After 1 h, the mixture was raised to rt and stirred overnight. It was then quenched with water and extracted twice with ether. The combined organic phase was washed with water and brine, dried over MgSO₄, and concentrated by rotary evaporation. The resulting crude oil was purified by silica gel chromatography (98:2 hexane/EtOAc) to give 8.3 g (56%) of **22** as an oil. ¹H NMR (300 MHz, CDCl₃) δ 3.70 (s, 3H), 3.44–3.36 (m+t, *J* = 6.0 Hz, 5H), 2.56–2.39 (m, 2H), 1.86–1.79 (m, 6H), 1.57–1.53 (m, 2H), 1.42–1.38 (m, 4H), 1.26 (m, 20H). ¹³C NMR (75 MHz, CDCl₃) δ 205.3, 170.3, 58.9, 52.2, 41.8, 33.9, 33.9, 32.7, 32.7, 29.3, 29.29, 29.28, 29.26, 29.25, 29.1, 28.7, 28.6, 28.12, 28.10, 23.4. IR 2987, 2854, 1745, 1716. HRMS (TOF MS ES⁺) calcd for C₂₃H₄₂Br₂O₃Na 547.1398; found 547.1407.

1,21-Dibromoheneicosan-11-one (23). To a mixture of **22** (2.04 g, 3.88 mmol) in 60 mL of diethyl ether was added 10 mL of satd NaOH (aq), and the mixture was refluxed overnight. The solvent was removed under reduced pressure, and the aqueous layer was extracted several times with ether. The organic layer was washed several times with water and dried over anhyd MgSO₄, and the solvent was removed under reduced pressure to yield 1.5 g of crude powder, which was purified by silica gel chromatography (97:3 hexane/EtOAc) to give 1.1 g (65%) as a white solid, mp 50–53 °C. ¹H NMR (300 MHz, CDCl₃) δ 3.40 (t, *J* = 6.0 Hz, 4H), 2.40–2.36 (m, 4H), 1.87–1.80 (m, 4H), 1.58–1.53 (m, 4H), 1.44–1.39 (m, 4H), 1.27 (m, 20H). ¹³C NMR (75 MHz, CDCl₃) δ 211.5, 42.7, 33.9, 32.8, 29.35, 29.32, 29.2, 28.7, 28.1, 23.8. HRMS (TOF MS ES⁺) calcd for C₂₁H₄₀Br₂ONa 489.1344; found 489.1347.

1,21-Dihydroxyheneicosan-11-one (24). A mixture of **23** (0.98 g, 2.09 mmol) and potassium acetate (616 mg, 6.28 mmol) in DMF (15 mL) was stirred for 2 h at 60 °C under nitrogen. Solvent was removed under reduced pressure to obtain crude diacetoxy ketone as a white solid. ¹H NMR (300 MHz, CDCl₃) δ 4.05 (t, J = 9.0, 4H), 2.38 (t, J = 6.0 Hz, 4H), 2.04 (s, 6H), 1.61–1.30 (m, 8H), 1.27 (m, 24H). ¹³C NMR (75 MHz, CDCl₃) δ 211.5, 171.1, 64.6, 42.7, 29.4, 29.37, 29.34, 29.2, 29.1, 28.5, 25.8, 23.8, 20.9. HRMS (TOF MS ES⁺) calcd for C₂₅H₄₆O₅Na 449.3243; found 449.3221.

Crude diacetoxy ketone was treated with KOH (700 mg) in methanol (20 mL) at 60 °C. The mixture was concentrated and extracted with dichloromethane. The extract was washed twice with water and dried over MgSO₄. Evaporation of solvent gave pure product as a white solid (0.40 g, 45%), which was used without purification. ¹H NMR (300 MHz, CDCl₃) δ 3.66 (t, *J* = 6.0 Hz, 4H), 2.40 (t, *J* = 9.0, 4H), 1.63–1.54 (m, 10H), 1.29 (m, 24H). ¹³C NMR (75 MHz, CDCl₃) δ 211.7, 63.0, 42.8, 32.7, 29.5, 29.3, 29.2, 25.7, 23.8. HRMS (TOF MS ES⁺) calcd for C₂₁H₄₂O₃Na 365.3032; found 365.3055.

11-Aminoheneicosane-1,21-diol (13d). To a solution of 24 (3.57 g, 10.4 mmol) in 150 mL of isopropyl alcohol were added NH₄OAc (8.03 g, 104 mmol), Na(BH₃)CN (655 mg, 10.4 mmol) and crushed 4 Å molecular sieves in one portion under nitrogen, and the mixture was stirred for 36 h at rt. The crushed molecular sieves were removed via vacuum filtration, and water was added to the filtrate, which was extracted twice with diethyl ether. The organic layer was washed with 1 M NaOH and brine and dried over anhyd MgSO₄ to obtain a brown oil after concentration, which was purified by silica gel chromatography (3:1:1 hexane/EtOAc/MeOH) to obtain 2.4 g of 13d

(69%) as a white solid, mp 99–100 °C. ¹H NMR (300 MHz, CDCl₃) δ 3.68–3.64 (m, 4H), 2.68 (m, 1H), 1.61–1.54 (m, 6H), 1.34–1.30 (m, 34H). ¹³C NMR (75 MHz, CDCl₃) δ 62.9, 51.3, 36.6, 32.7, 29.7, 29.6, 29.4, 29.38, 29.32, 25.8, 25.7. HRMS (TOF MS ES⁺) calcd for C₂₁H₄₅NO₂Na 366.3348; found 366.3371.

N,*N*′-Bis(1,21-dihydroxyheneicos-11-yl)perylene-3,4,9,10bis(dicarboximide) (6d). A mixture of 1.14 g (2.91 mmol) of 5, 2.2 g (6.4 mmol) of 13d, 5 g of imidazole, and a catalytic amount of zinc acetate was heated with stirring at 130 °C for 6 h. The mixture was then cooled, dissolved in CHCl₃, and directly purified by column chromatography on silica gel (CHCl₃) to give 1.6 g (55%) of a deep red, waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.68–8.58 (m, 8H), 5.19 (m, 2H), 3.61 (t, *J* = 6.0 Hz, 8H), 2.30–2.23 (m, 4H), 1.92–1.83 (m, 4H), 1.55–1.48 (m, 12H), 1.30–1.23 (m, 56H). ¹³C NMR (75 MHz, CDCl₃) δ 164.6, 163.5, 134.5, 131.9, 131.1, 129.5, 126.4, 123.8, 123.0, 63.0, 54.7, 32.7, 32.3, 29.49, 29.46, 29.42, 29.3, 26.9, 25.6. HRMS (TOF MS ES⁺) calcd for C₆₆H₉₄N₂O₈Na 1065.6908; found 1065.6935.

N-(1,21-Dihydroxyheneicos-11-yl)-3,4,9,10-perylenetetracarboxylic Acid 3,4-Anhydride 9,10-Imide (7d). A mixture of 600 mg (0.57 mmol) of 6d, 97.0 mg (1.75 mmol) of 85% KOH pellets, and 15 mL of t-BuOH was brought to reflux. After 1 h an additional 4 pellets of KOH were added, and heating was continued for an additional 1 h. The reaction was monitored with TLC and stopped after appropriate conversion had occurred. The mixture was poured, with stirring, into a mixture of AcOH and 2 N HCl. The resulting solution was extracted with chloroform, washed twice with water and brine, and purified by silica gel chromatography using 19:1 CHCl₃/ MeOH to obtain 178 mg (43%) of monoimide 7d as a waxy solid. ¹H NMR (500 MHz, CDCl₃) δ 8.74-8.68 (m, 8H), 5.24-5.17 (m, 1H), 3.64-3.62 (m, 4H), 2.28-2.25 (m, 2H), 1.89-1.86 (m, 2H), 1.58-1.35 (m, 6H), 1.34–1.25 (m, 28H). 13 C NMR (125 MHz, CDCl₃) δ 164.4, 163.3, 159.9, 135.2, 133.6, 133.5, 131.8, 131.1, 129.5, 126.8, 126.5, 124.6, 123.9, 123.1, 119.0, 63.0, 54.9, 32.7, 32.3, 29.48, 29.45, 29.42, 29.3, 26.9, 25.6. HRMS (TOF MS ES⁺) calcd for C₄₅H₅₁NO₇Na 740.3563; found 740.3589.

N-(1,21-Diacetylthioheneicos-11-yl)- 3,4,9,10-perylenetetracarboxylic Acid 3,4-Anhydride 9,10-Imide (7e). *Method A*. Diol monoimide 7d (50.2 mg, 0.070 mmol) was dissolved in 4 mL of pyridine, and the reaction mixture was cooled to 0 °C. Tosyl chloride (29.3 mg, 0.154 mmol) dissolved in pyridine was added dropwise at 0 °C, and the reaction mixture was stirred overnight at rt. When TLC showed no starting material left, the reaction mixture was diluted with CH_2Cl_2 , washed with water and then brine, dried over anhyd MgSO₄, and concentrated by rotary evaporation to obtain 60.7 mg of crude ditosylate monoimide that was used further without purification.

Ditosylate monoimide (50.3 mg, 0.049 mmol) and KSAc (13.9 mg, 0.123 mmol) were dissolved in 5 mL of DMF and stirred overnight at 120 °C. The solvent was removed under reduced pressure, and the residue was dissolved in CH_2Cl_2 , washed with water and then brine, and dried over anhyd MgSO₄. After rotary evaporation, the residue was purified by silica gel chromatography (1:99 MeOH/CHCl₃) to yield 22 mg (54%) of pure 7e as a waxy solid.

Method B. Diene monoimide 7h (417 mg, 0.61 mmol) was dissolved in 20 mL of toluene. To this solution were added thiolacetic acid (0.17 mL, 2.4 mmol) and a catalytic amount of AIBN. The solution was then refluxed for 1 h. The reaction was quenched by addition of 1 M NaHCO3 and extracted with CHCl3. The organic layer was washed three times with 1 M NaHCO₃ and then brine, concentrated under reduced pressure, and purified by silica gel chromatography (1:99 MeOH/CHCl₃) to yield 428 mg (84%) of pure 7e as a waxy solid. ¹H NMR (500 MHz, CDCl₃) δ 8.78–8.52 (m, 8H), 5.22-5.16 (m, 1H), 2.82 (t, J = 10.0 Hz, 4H), 2.30 (s+m, 6H + 2H),1.89 (m, 2H), 1.53–1.50 (m, 4H) 1.35–1.22 (m, 28H). ¹³C NMR (125 MHz, CDCl₃) δ 195.9, 164.2, 163.1, 159.7, 136.1, 133.3, 131.8, 131.6, 131.1, 129.3, 129.5, 126.3, 124.7, 124.0, 123.8, 123.0, 118.8, 54.9, 32.3, 30.6, 29.49, 29.47, 29.44, 29.3, 29.1, 29.0, 28.7, 26.9. HRMS (TOF MS ES⁺) calcd for C₄₉H₅₅NO₇ S₂Na 856.3318; found 856.3337. N-(1,21-Dimercaptoheneicos-11-yl)-3,4,9,10-perylenetetra-

carboxylic Acid 3,4-Anhydride 9,10-Imide (7f). Dithioacetate

monoimide 7e (108 mg, 0.129 mmol) was deprotected by addition of acetyl chloride (2 mL) at 0 °C to a solution in methanol (2 mL) and dichloromethane (10 mL). After overnight stirring at rt the solvent was removed in vacuum, and the residue was dissolved in 20 mL of CHCl₃. The solution was washed with 5% NaHCO₃, dried over MgSO₄, concentrated under reduced pressure, and purified by silica gel chromatography (1:99 MeOH/CHCl₃) to yield 92 mg (95%) of pure 7f as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.75–8.69 (m, 8H), 5.23–5.18 (m, 1H), 2.53–2.48 (q, *J* = 9.0 Hz 4H), 2.30–2.24 (m, 2H), 1.87–1.82 (m, 2H), 1.59–1.55 (m, 4H), 1.36–1.24 (m, 30H). ¹³C NMR (75 MHz, CDCl₃) δ 164.2, 163.1, 159.6, 136.0, 133.3, 133.2, 131.7, 131.5, 131.0, 130.0, 129.2, 126.4,126.2, 124.7, 123.8, 122.9, 122.0, 121.6, 118.7, 54.9, 34.0, 32.3, 29.5, 29.49, 29.48, 29.43, 29.0, 28.3, 26.9, 24.6. HRMS (TOF MS ES⁺) calcd for C₄₅H₅₁NS₂O₅Na 772.3106; found 772.3124.

N-(1,21-Bis(methyldisulfanyl)heneicos-11-yl)-3,4,9,10-perylenetetracarboxylic Acid 3,4-Anhydride 9,10-Imide (7g). To a solution of 7f (47 mg, 0.063 mmol) in CH22Cl2 was added methyl methanethiosulfonate (MMTS) (17.7 µL, 0.188 mmol) and triethylamine (26.2 μ L, 0.188 mmol) at room temperature. After 5 h of stirring, the reaction mixture was washed with 5% HCl, water, and brine, dried over MgSO4, concentrated under reduced pressure, and purified by silica gel chromatography (1:99 MeOH/CHCl₃) to give 92 mg (95%) of pure 7g as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.67–8.50 (m, 8H), 5.22–5.16 (m, 1H), 2.66 (t, J = 9.0 Hz, 4H), 2.38 (s, 6H), 2.32-2.26 (m, 2H), 2.24-2.21 (m, 2H), 1.94-1.85 (m, 4H), 1.69-1.23 (m, 28H). ¹³C NMR (75 MHz, CDCl₃) δ 164.3, 163.1, 159.7, 136.1, 133.3, 131.8, 131.5, 131.1, 129.3, 126.4, 126.2, 123.8, 123.0, 118.7, 54.9, 38.3, 32.3, 29.5, 29.47, 29.45, 29.41, 29.1, 28.4, 26.9, 23.3. HRMS (TOF MS ES^+) calcd for $C_{47}H_{55}NO_5S_4Na$ 864.2861; found 864.2869.

Methyl 2-(Non-8-enyl)-3-oxotridec-12-enoate (26). A solution of 11-undecenoic acid (25) (45.7 g, 248 mmol) and concd H_2SO_4 (5 mL) in MeOH (600 mL) was refluxed for 6 h. After most of the solvent was removed under reduced pressure, water was added, and the mixture was extracted with ether. The ether layer was washed with 5% NaHCO₃, water, and brine, dried over anhyd MgSO₄, and concentrated by rotary evaporation to yield 45 g (92%) of crude methyl undecanoate, which was used without purification in the next step. ¹H NMR was in agreement with the literature.⁵²

TiCl₄ (6.11 g, 32.2 mmol) in toluene (50 mL) was added to a stirred solution of methyl undecanoate (5.16 g, 17.9 mmol) and Bu₃N (4.98 g, 26.9 mmol) in toluene at 0-5 °C. After 1 h, the mixture was raised to rt and stirred overnight. It was then quenched with water and extracted twice with ether. The combined organic phase was washed with water and brine, dried over MgSO4, and concentrated by rotary evaporation. The resulting crude oil was purified by silica gel chromatography (98:2 hexane/EtOAc) to give 7.0 g (74%) of 26 as an oil. ¹H NMR (300 MHz, CDCl₃) δ 5.85–5.76 (m, 2H), 5.03–4.91 (m, 4H), 3.72 (s, 3H), 3.46-3.41 (m, 1H), 2.54-2.46 (m, 2H), 2.07-2.00 (m, 4H), 1.84–1.60 (m, 2H), 1.57–1.32 (m, 2H), 1.28–1.24 (m, 20H). ¹³C NMR (75 MHz, CDCl₃) δ 205.4, 170.4, 139.1, 139.0, 114.16, 114.13, 58.9, 52.2, 41.8, 33.78, 33.75, 29.35, 29.31, 29.2, 29.1, 29.04, 29.00, 28.9, 28.88, 28.83, 28.2, 27.4, 23.4. IR 2927, 2855, 1747, 1717. HRMS (TOF MS ES⁺) calcd for $C_{23}H_{40}O_3Na$ 387.2875; found 387.2898.

Heneicosa-1,20-dien-11-one (27). To a solution of 26 (4.0 g, 7.6 mmol) in 100 mL of THF was added 20 mL of satd NaOH (aq), and the mixture was refluxed overnight. The solvent was removed under reduced pressure, and the aqueous layer was extracted several times with ether. The organic layer was washed several times with water and dried over anhyd MgSO₄, and the solvent was removed under reduced pressure to yield 3.0 g of crude powder, which was purified by silica gel chromatography (97:3 hexane/EtOAc) to give 2.83 g (84%) of 27, mp 55–56 °C. ¹H NMR (500 MHz, CDCl₃) δ 5.87–5.81 (m, 2H), 5.03–4.94 (m, 4H), 2.43–2.39 (m, 4H), 2.08–2.04 (m, 4H), 1.60–1.58 (m+s, 6H + H₂O), 1.39–1.38 (m, 4H), 1.30 (m, 14H). ¹³C NMR (125 MHz, CDCl₃) δ 211.8, 139.2, 114.1, 42.8, 33.8, 29.38, 29.33, 29.2, 29.0, 28.9, 23.8. IR 1698, 994, 915. HRMS (TOF MS ES⁺) calcd for C₂₁H₃₈ONa 329.2820; found 329.2790.

Heneicosa-1,20-dien-11-amine (13h). To a solution of 27 (5.06 g, 16.5 mmol) in 150 mL of isopropyl alcohol were added NH₄OAc (12.7 g, 165 mmol), Na(BH₃)CN (1.03g, 16.5 mmol), and crushed 4 Å molecular sieves in one portion under nitrogen, and the mixture was stirred for 36 h at rt. The crushed molecular sieves were removed via vacuum filtration, and deionized water was added to the filtrate, which was extracted twice with diethyl ether. The organic layer was washed with 1 M NaOH and brine and dried over anhyd MgSO4 to obtain a brown oil after concentration, which was purified by silica gel chromatography (3:1:1 hexane/EtOAc/MeOH) to obtain 3.7 g of 13h (73%) as a white solid. ¹H NMR (300 MHz, CDCl₃) δ 5.89–5.76 (m, 2H), 5.04-4.92 (m, 4H), 2.69-2.67 (m, 1H), 2.09-2.02 (m, 4H), 1.41-1.38 (m, 10H), 1.36-1.21 (m, 20H). ¹³C NMR (75 MHz, CDCl₃) & 139.1, 114.1, 51.2, 37.3, 33.8, 29.7, 29.5, 29.4, 29.1, 28.9, 26.0. HRMS (TOF MS ES⁺) calcd for C₂₁H₄₁NNa 330.3137; found 330.3166.

N,*N*′-**Di**(1,20-heneicosadien-11-yl)perylene-3,4,9,10-bis-(dicarboximide) (6h). A mixture of 3.07 g (7.83 mmol) of 5, 5.06 g (16.4 mmol) of 13h, 15 g of imidazole, and a catalytic amount of zinc acetate was heated with stirring at 160 °C for 2 h. The mixture was then cooled, dissolved in CHCl₃, and directly purified by column chromatography on silica gel (CHCl₃) to give 3.9 g (74%) of a deep red, waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.70–8.64 (m, 8H), 5.84–5.70 (m, 4H) 5.19 (m, 2H), 4.98–4.87 (m, 8H), 2.25 (m, 4H), 2.03–1.95 (m, 8H), 1.87 (m, 4H), 1.32–1.23 (m, 48H). ¹³C NMR (125 MHz, CDCl₃) δ 164.6, 163.5, 139.2, 134.4, 131.8, 131.1, 129.5, 126.4, 123.9, 123.0, 114.0, 54.7, 33.7, 32.3, 29.5, 29.4, 29.08, 29.01, 28.8, 26.9. HRMS (TOF MS ES⁺) calcd for C₆₆H₈₆N₂O₄Na 993.6485; found 993.6491.

N-(1,20-Heneicosadien-11-yl)-3,4,9,10-perylenetetracarboxylic Acid 3,4-Anhydride-9,10 Imide (7h). A mixture of 1.24 g (1.27 mmol) of 6h, 215 mg (3.83 mmol) of 85% KOH pellets, and 50 mL of t-BuOH was brought to reflux. After 1 h an additional 10 pellets of KOH were added, and heating was continued for an additional 1 h. The reaction was monitored with TLC and stopped after appropriate conversion had occurred. The mixture was poured with stirring into a mixture of 70 mL of AcOH and 40 mL of 2 N HCl. The resulting solution was extracted with chloroform, which was washed twice with water and once with brine, concentrated, and purified by silica gel chromatography using CHCl₃ to obtain 520 mg (59%) of monoimide 7h as a waxy solid. ¹H NMR (500 MHz, CDCl₃) δ 8.66–8.46 (m, 8H), 5.84-5.70 (m, 2H), 5.22-5.13 (m, 1H), 4.98-4.87 (m, 4H), 2.27-2.01 (m, 2H), 1.98-1.89 (m+m, 2H+4H), 1.37-1.25 (m, 24H). $^{13}\mathrm{C}$ NMR (125 MHz, $\mathrm{CDCl}_3)$ δ 164.3, 163.2, 159.8, 139.1, 136.2, 133.4, 131.7, 129.4, 126.6, 126.4, 123.8, 123.0, 118.9, 114.0, 54.8, 33.7, 32.2, 29.46, 29.42, 29.3, 29.06, 29.03, 28.8, 26.9. HRMS (TOF MS ES⁺) calcd for C₄₅H₄₇NO₅Na 704.3352; found 704.3381.

General Procedure for Bisimide D- σ -A Molecules with a PEG Swallowtail (R = b). Perylene monoimide monoanhydride with a PEG swallowtail (7b) and a suitable donor amine were added to pyridine and stirred under reflux for 12 h. Then the reaction mixture was added to excess 5% HCl and extracted with CHCl₃. The organic layer was then washed with an excess of 10% K₂CO₃, dried over anhyd Na₂SO₄, and rotary evaporated to give crude product, which was purified by silica gel chromatography. Pyrenemethylamine (1-NH₂) was prepared by treating pyrenemethylammonium chloride (Aldrich) with KOH (aq) and MeOH.

N-(2,5,8,12,15,18-Hexaoxanonadec-10-yl)-*N'*-(1pyrenylmethyl)perylene-3,4,9,10-bis(dicarboximide) (1b). Prepared from 7b (98.0 mg, 0.146 mmol) and 1-pyrenylmethylamine (1-NH₂, 43.0 mg, 0.186 mmol). Elution with 99:1 CHCl₃/MeOH gave 25 mg (19%) of 1b as a waxy solid. ¹H NMR (500 MHz, CDCl₃) δ 8.64–8.14 (m, 5H), 8.11–8.07 (d, 4H), 8.05–7.82 (m, 8H), 6.11 (s, 2H), 5.74–5.72 (m, 1H), 4.27–4.21 (m, 2H), 4.04–3.99 (m, 2H), 3.77–3.57 (m, 12H), 3.46–3.43 (m, 4H), 3.30 (s, 6H). ¹³C NMR (75 MHz, CDCl₃) δ 163.5, 163.3, 133.8, 133.4, 131.0, 130.6, 130.2, 129.9, 128.6, 128.5, 128.3, 127.5, 126.8, 126.7, 125.5, 125.4, 125.3, 125.2, 124.9, 124.8, 124.5, 124.2, 124.0, 122.8, 122.4, 122.2, 71.8, 70.5, 70.4, 70.3, 69.4, 58.9, 52.1, 41.2. IR (KBr) 1692 (s), 1656 (s), 1595 (s), 1341 (s), 1252 (m), 1110 (s) cm⁻¹. HRMS (TOF MS ES⁺) calcd for $C_{54}H_{46}N_2O_{10}Na$ 905.3050; found 905.3077.

N-(2,5,8,12,15,18-Hexaoxanonadec-10-yl)-*N'*-(4-[1-pyrenyl]butyl)perylene-3,4,9,10-bis(dicarboximide) (2b). Prepared from 7b (102 mg, 0.152 mmol) and 4-pyrenylbutylamine (2-NH₂, Toronto Research Chemicals, 52.0 mg, 0.190 mmol). Elution with 99:1 CHCl₃/ MeOH gave 26 mg (18%) of 2b as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.37–8.21 (m, 4H), 8.08–7.62 (m, 13H), 5.78–5.72 (m, 1H), 4.31–4.20 (m, 4H), 4.08–4.02 (m, 2H), 3.85–3.59 (m, 12H), 3.47 (t, *J* = 6.0 Hz, 4H), 3.31 (s, 6H), 3.14 (m, 2H), 1.95 (m, 4H). ¹³C NMR (75 MHz, CDCl₃) δ163.7, 163.0, 136.1, 133.8, 133.7, 130.9, 130.7, 130.3, 129.3, 128.4, 128.0, 127.3, 127.1, 126.8, 126.1, 125.4, 124.5, 124.4, 124.3, 123.0, 122.4, 122.3, 71.8, 70.5, 70.4, 70.3, 69.4, 58.9, 52.0, 40.2, 33.3, 28.8, 27.8. IR (KBr) 1692 (s), 1655 (s), 1595 (m), 1342 (s), 1253 (m), 1109 (s) cm⁻¹. HRMS (TOF MS ES⁺) calcd for C₅₇H₅₂N₂O₁₀Na 947.3520; found 947.3517.

N-(2-Ferrocenylethyl)-*N*'-(2,5,8,12,15,18-hexaoxanonadec-10-yl)perylene-3,4,9,10-bis(dicarboximide) (3b). Prepared from 7b (119 mg, 0.178 mmol) and 3-ferrocenylethylamine¹ (3-NH₂, 51 mg, 0.22 mmol). Elution with 99.4:0.6 CHCl₃/MeOH gave 47 mg (24%) of 3b as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.59– 8.51 (m, 4H), 8.44–8.39 (m, 4H), 5.77–5.68 (q, 1H), 4.37 (m, 2H), 4.22 (m, 9H), 4.10 (s, 2H), 3.65 (m, 2H), 3.63–5.56 (m, 12H), 3.45– 3.44 (m, 4H), 3.28 (s, 6H), 2.82–2.77 (m, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 163.6, 163.1, 134.4, 134.1, 131.2, 129.3, 129.1, 126.2, 126.0, 123.4, 123.0, 122.96, 122.92, 85.1, 71.8, 70.5, 70.4, 70.3, 68.6, 58.9, 52.1, 41.2, 27.7. IR (KBr) 1696 (s), 1655 (s), 1596 (m), 1342 (s), 1249 (m), 1105 (s) cm⁻¹. HRMS (TOF MS ES⁺) calcd for C₄₉H₄₈FeN₂O₁₀Na 903.2557; found 903.2574.

N-(3-[2, 5-Bis (dimethylamino)phenyl]propyl)-*N'*-(2,5,8,12,15,18-hexaoxanonadec-10-yl)perylene-4,5,9,10-bis-(dicarboximide) (4b). Prepared from 7b (126 mg, 0.188 mmol) and 3-tetramethylphenylenediaminylpropylamine¹ (4-NH₂, 70.0 mg, 0.316 mmol). Elution with 49:1 CHCl₃/MeOH gave 21 mg (13%) of 4b as a waxy solid and 50 mg (31%) of an additional fraction containing a trace impurity by TLC. ¹H NMR (500 MHz, CDCl₃) δ 8.57–8.51 (m, 4H), 8.42–8.35 (m, 4H), 7.05–7.03 (d, 1H), 6.71 (s, 1H), 6.56–6.55 (d, 1H), 5.75–5.69 (m, 1H), 4.32–4.30 (m, 2H), 4.20–4.23 (m, 2H), 3.97–4.01 (m, 2H), 3.64–3.57 (m, 12H), 3.44–3.42 (m, 4H), 3.28 (s, 6H), 2.89 (s, 8H), 2.62 (s, 6H) 2.19–2.13 (m, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 163.6, 163.0, 147.5, 143.1, 137.4, 134.0, 131.0, 129.2, 128.9, 125.9, 125.8, 123.2, 123.1, 122.8, 122.7, 120.6, 114.2, 111.3, 71.8, 70.5, 70.4, 70.3, 69.3, 58.9, 52.1, 45.8, 41.1, 40.9, 29.7, 28.6. IR (KBr) 1698 (s), 1655 (s), 1596 (m), 1343 (s), 1107 (s) cm⁻¹. HRMS (TOF MS ES⁺) calcd for C₅₀H₅₆N₄O₁₀Na 895.3894; found 895.3890.

General Procedure for Bisimides D- σ -A Molecules with a Mixed Swallowtail (R = c). Perylene monoimide monoanhydride with a mixed alkyl-PEG swallowtail 7c and a donor amine were dissolved in toluene and refluxed for 12 h to give the target nonsymmetric bisimides 1c-4c. These were purified by neutral Al₂O₃ column chromatography (CHCl₃) followed by silica gel column chromatography (99:1 CHCl₃/MeOH).

N-(1-Pyrenylmethyl)-*N'*-(2,5,15,18-tetraoxanonadec-10-yl)perylene-3,4,9,10-bis(dicarboximide) (1c). Pyrenylmethylamine 1-NH₂ (35 mg, 0.15 mmol) and 7c (70.0 mg, 0.105 mmol) gave 30 mg (27%) of 1c as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.41– 8.35 (d, *J* = 9.4 Hz, 3H), 8.16–8.13 (d, *J* = 7.9 Hz, 2H), 7.95 (d, *J* = 7.9 Hz, 2H), 7.74 (m, 8H), 7.54 (m, 2H), 5.86 (s, 2H), 5.23 (m, 1H), 3.47 (m, 12H), 3.31 (s, 6H), 2.34 (m, 2H), 1.99 (m, 2H), 1.68 (m, 4H), 1.46 (m, 4H). ¹³C NMR (75 MHz, CDCl₃) δ 164.3, 163.2, 133.6, 133.3, 130.7, 130.6, 130.18, 130.15, 129.9, 128.7, 128.3, 127.4, 126.7, 125.6, 125.5, 125.2, 125.1, 124.8, 124.7, 124.4, 124.2, 123.9, 122.8, 122.6, 122.4, 122.2, 122.0, 71.8, 71.2, 69.8, 58.9, 54.5, 41.1, 32.2, 29.5, 23.6. IR (KBr) 1692 (s), 1655 (s), 1595 (s) cm⁻¹. HRMS (TOF MS ES⁺) calcd for C₅₆H₅₀N₂O₈Na 901.3465; found 901.3491.

N-[4-(1-Pyrenyl)butyl]-*N*'-(2,5,15,18-tetraoxanonadec-10-yl)perylene-3,4,9,10-bis(dicarboximide) (2c). Pyrenebutylamine (2-NH₂, 15 mg, 0.056 mmol) and 7c (25 mg, 0.038 mmol) gave 7.5 mg (22%) of 2c as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.64 (m, 2H), 8.45 (dxd, *J* = 9.0 Hz, 3.0 Hz, 4H), 8.34 (d, *J* = 9.0 Hz, 2H), 8.14 (d, *J* = 9.0 Hz, 1H), 8.00–7.40 (m, 8H), 5.24 (m, 1H), 4.25 (s, 2H), 3.54–3.46 (m, 12H), 3.33 (s, 6H), 2.32–2.34 (m, 2H), 1.94–2.02 (m +m, 2H + 4H), 1.60–1.69 (m, 6H + H₂O), 1.44 (m, 4H). ¹³C NMR (75 MHz, CDCl₃) δ 164.6, 163.2, 143.4, 136.2, 134.2, 131.0, 130.5, 129.5, 128.3, 127.5, 127.3, 126.9, 126.2, 125.9, 125.5, 124.6, 124.5, 123.2, 122.8, 122.7, 122.6, 71.8, 71.2, 69.9, 59.0, 54.5, 40.3, 33.4, 32.1, 29.4, 28.6, 27.6, 23.5. HRMS (TOF MS ES⁺) calcd for C₅₉H₅₆N₂O₈Na 943.3934; found 943.3951.

N-(2-Ferrocenylethyl)-*N*'-(2,5,15,18-tetraoxanonadec-10-yl)perylene-3,4,9,10-bis(dicarboximide) (3c). Ferrocenylethylamine 3-NH₂ (62 mg, 0.28 mmol) and 7c (80 mg, 0.12 mmol) in 5 mL of toluene gave 21 mg (20%) of 3c as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.66–8.55 (m, 8H), 5.21 (m, 1H), 4.43–4.37 (m, 2H), 4.22 (s, 7H), 4.12 (s, 2H), 3.51–3.43 (m, 12H), 3.32 (s, 6H), 2.81–2.76 (m, 2H), 2.32–2.29 (m, 2H), 1.95–1.91 (m, 2H), 1.71–1.66 (m, 4H), 1.65–1.40 (m, 4H). ¹³C NMR (75 MHz, CDCl₃) δ 163.4, 163.1, 134.6, 134.2, 131.3, 129.4, 126.4, 123.1, 123.06, 123.00, 85.0, 71.8, 71.2, 69.9, 68.6, 68.1, 67.4, 59.0, 54.5, 41.2, 32.1, 29.4, 27.7, 23.5. IR (KBr) 1692 (s), 1656 (s), 1596 (s) cm⁻¹. HRMS (TOF MS ES⁺) calcd for C₅₁H₅₂FeN₂O₈Na 899.2972; found 899.2991.

N-(3-(2,5-Bis(dimethylamino)phenyl]propyl)-*N*'-(2,5,15,18tetraoxanonadec-10-yl)perylene-4,5,9,10-bis(dicarboximide) (4c). TMPDA-propanamine 4-NH₂ (80.0 mg, 0.361 mmol) and 7c (120 mg, 0.180 mmol) gave 44 mg (28%) of 4c as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.66–8.56 (m, 8H), 7.06–7.04 (d, *J* = 6.0 Hz, 1H), 6.72–6.71 (d, *J* = 3.0 Hz, 1H), 6.58–6.55 (d × d, *J* = 10 Hz, 5.0 Hz, 1H), 5.23–5.21 (m, 1H), 4.35–4.32 (m, 2H), 3.54–3.45 (m, 12H), 3.33 (s, 6H), 2.90–2.89 (m, 6 + 2H), 2.63 (s, 6H), 2.31 (m, 2H), 2.14 (m, 2H), 1.92 (m, 2H), 1.67 (4H+H₂O), 1.38 (m, 4H). ¹³C NMR (75 MHz, CDCl₃) δ 164.5, 163.2, 147.5, 137.4, 131.2, 129.3, 126.4, 123.3, 122.98, 122.95, 120.6, 114.2, 111.3, 71.9, 71.2, 69.9, 58.9, 54.5, 45.8, 41.1, 40.8, 32.1, 29.4, 28.6, 23.5. IR (KBr) 1695 (s), 1655 (s), 1595 (s) cm⁻¹. HRMS (TOF MS ES⁺) calcd for C₅₂H₆₀N₄O₈Na 891.4309; found 891.4333.

General Procedure for Bisimide D- σ -A Molecules with a Dithioacetyl Swallowtail (R = e). A mixture of the perylene monoimide monoanhydride with a dithioacetylalkyl swallowtail (7e) and a 2- to 4-fold molar excess of the appropriate amine was refluxed in toluene for 1–2 h. The reaction mixture was then cooled, washed with 5% HCl followed by 10% K₂CO₃, dried over MgSO₄, and concentrated by rotary evaporation. Crude products were purified by chromatography on silica gel (1:99 MeOH/CHCl₃) to give red solids. Melting points were difficult to obtain due to the deep color and waxy nature of the materials.

N-(1,21-Diacetylthioheneicos-11-yl)-*N*'-([1-pyrenyl]methyl)perylene-3,4,9,10-bis(dicarboximide) (1e). 1-NH₂ (20.7 mg, 0.090 mmol) and 7h (47.0 mg, 0.045 mmol) gave 33 mg (88%) of pure 1e as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.68–8.61 (m, SH), 8.53–8.47 (m, 4H), 8.19–8.04 (m, SH), 7.97–7.90 (m, 3H), 6.12 (s, 2H), 5.20 (m, 1H), 2.81 (t, *J* = 6.0 Hz, 4H), 2.30–2.25 (s+m, 6H + 2H), 1.88–1.85 (m, 2H), 1.53–1.45 (m, 4H), 1.30–1.19 (m, 28H). ¹³C NMR (75 MHz, CDCl₃) δ 196.2, 164.8, 163.7, 134.7, 134.1, 131.6, 131.3, 130.8, 130.8, 130.3, 129.5, 129.3, 128.9, 127.9, 127.4, 127.3, 126.2, 126.2, 126.0, 125.3, 125.2, 124.9, 124.8, 124.7, 124.1, 123.3, 123.2, 122.9, 55.0, 41.6, 32.6, 30.8, 29.79, 29.75, 29.72, 29.6, 29.38, 29.31, 29.2, 29.0, 27.2. HRMS (TOF MS ES⁺) calcd for C₆₆H₆₆N₂O₆ S₂Na 1069.4260; found 1069.4248.

N-(1,21-Diacetylthioheneicos-11-yl)-*N*'-(4-[1-pyrenyl]butyl)perylene-3,4,9,10-bis(dicarboximide) (2e). 2-NH₂ (43.30 mg, 0.158 mmol) and 7e (66.00 mg, 0.079 mmol) gave 2e in 82% yield as a waxy solid. ¹H NMR (500 MHz, CDCl₃) δ 8.46–7.52 (m, 17H), 5.26–5.18 (m, 1H), 4.17–4.12 (m, 2H), 3.12–3.07 (m, 2H), 2.81 (t, *J* = 9.0 Hz, 4H), 2.32–2.27 (s+m 6H + 2H), 1.94–1.91 (m, 4H), 1.55– 1.45 (m, 6H), 1.37–1.23 (m, 28H). ¹³C NMR (75 MHz, CDCl₃) δ 196.0, 164.4, 162.9, 136.1, 133.6, 130.8, 130.5, 130.3, 129.2, 128.9, 128.3, 127.9, 127.2, 127.0, 126.7, 126.0, 125.4, 125.3, 124.5, 124.38, 124.32, 124.2, 122.9, 122.4, 122.2, 122.1, 54.6, 40.1, 33.3, 32.3, 30.5, 29.55, 29.52, 29.48, 29.43, 29.3, 29.07, 29.05, 28.9, 28.7, 27.0. HRMS (TOF MS ES⁺) calcd for C₆₉H₇₂N₂O₆S₂Na 1111.4729; found 1111.4746. *N*-(1,21-Diacetylthioheneicos-11-yl)-*N*'-(2-ferrocenylethyl)perylene-3,4,9,10-bis(dicarboximide) (3e). 3-NH₂ (27.50 mg, 0.120 mmol) and 7e (50.1 mg, 0.06 mmol) gave 3e (54 mg) in 86% yield as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.69–8.53 (m, 8H), 5.23–5.18 (m, 1H), 4.40 (t, *J* = 6.0 Hz, 2H), 4.23 (s,7H), 4.16 (s, 2H), 2.85–2.76 (m, 6H), 2.31–2.25 (m, 6H + 2H), 1.92–1.85 (m, 2H), 1.60–1.47 (m, 4H), 1.32–1.22 (m, 28H). ¹³C NMR (75 MHz, CDCl₃) δ 196.0, 164.5, 163.1, 134.6, 134.2, 131.2, 129.4, 129.2, 126.3, 126.2, 123.0, 122.9, 85.0, 68.6, 68.1, 67.4, 54.7, 41.2, 32.3, 30.6, 29.5, 29.46, 29.44, 29.3, 29.1, 29.0, 28.7, 27.7, 26.9. HRMS (TOF MS ES⁺) calcd for C₆₁H₆₈FeN₂O₆S₂Na 1067.3768; found 1067.3789.

N-(1,21-Diacetylthioheneicos-11-yl)-*N*'-(3-[2-(*N*',*N*',*N*",*N*"-tetramethyl-1,4-benzenediaminyl)propyl)perylene-3,4,9,10-bis(dicarboximide) (4e). 4-NH₂ (7.96 mg, 0.036 mmol) and 7e (30.0 mg, 0.036 mmol) gave 4e (26 mg) in 71% yield as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.53–8.46 (m, 2H), 8.31–8.12 (m, 6H), 7.03–7.00 (d, *J* = 6.0 Hz, 1H), 6.70 (d, *J* = 3.0 Hz, 1H), 6.54–6.50 (dxd, *J* = 9.0 Hz, 3.0 Hz, 1H), 5.21–5.14 (m, 1H), 4.25–4.20 (m, 2H), 2.88–2.78 (m, 10H), 2.60 (s, 6H), 2.29–2.28 (m, 8H), 2.16–2.05 (m, 2H), 1.94–1.90 (m, 2H), 1.54–1.42 (m, 6H), 1.37–1.22 (m, 28H). ¹³C NMR (75 MHz, CDCl₃) δ 195.8, 164.1, 162.6, 147.3, 143.0, 137.3, 133.7, 130.5, 129.0, 125.6, 122.7, 122.4, 120.4, 114.0, 111.1, 54.6, 45.7, 41.0, 33.9, 32.2, 30.5, 29.47, 29.43, 29.40, 29.37, 29.33, 29.0, 28.7, 28.4, 26.9. HRMS (TOF MS ES⁺) calcd for C₆₂H₇₆ N₄O₆S₂Na 1059.5104; found 1059.5125.

N-(1,21-Dimercaptoheneicos-11-yl)-*N*'-([1-pyrenyl]methyl)perylene-3,4,9,10-bis(dicarboximide) (1f). 1-NH₂ (18.50 mg, 0.080 mmol) and 7f (30.00 mg, 0.040 mmol) gave 1f (34 mg) in 89% yield as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.54–8.49 (m, 3H), 8.39 (d, *J* = 6.0 Hz, 2H), 8.26 (d, *J* = 9.0, 2H), 8.18 (d, *J* = 9.0, 2H), 8.03–7.70 (m, 8H), 5.98 (s, 2H), 5.20 (m, 1H), 2.50–2.43 (q, *J* = 6.0 Hz, 4H), 2.33–2.25 (m, 2H), 1.93–1.88 (m, 2H), 1.56– 1.48 (m, 4H), 1.31–1.18 (m, 30H). ¹³C NMR (75 MHz, CDCl₃) δ 163.4, 134.2, 133.7, 131.3, 130.9, 130.49, 130.44, 130.0, 129.1, 128.9, 128.6, 127.6, 127.0, 125.8, 125.6, 125.0, 124.9, 124.58, 124.54, 124.3, 123.0, 122.8, 122.5, 54.7, 41.3, 33.9, 32.3, 29.55, 29.50, 29.47, 29.40, 28.9, 28.3, 27.0, 24.6. HRMS (TOF MS ES⁺) calcd for C₆₂H₆₂ N₂O₄ S₂Na 985.4048; found 985.4070.

N-(1,21-Dimercaptoheneicos-11-yl)-*N'*-(2-ferrocenylethyl)perylene-3,4,9,10-bis(dicarboximide) (3f). 3-NH₂ (18.30 mg, 0.080 mmol) and 7f (30.0 mg, 0.04 mg) gave 3f (33 mg) in 87% yield as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.69–8.60 (m, 8H), 5.20 (m, 1H), 4.43–4.38 (m, 2H), 4.24–4.21 (m, 7H), 4.11 (s, 2H), 2.79 (t, J = 9.0 Hz, 2H), 2.52–2.44 (q, J = 6.0 Hz, 4H), 2.33– 2.20 (m, 2H), 1.93–1.83 (m, 2H), 1.58–1.51 (m, 4H + H₂O), 1.32– 1.22 (m, 30H). ¹³C NMR (75 MHz, CDCl₃) δ 163.2, 134.7, 134.3, 131.8, 131.4, 131.1, 129.5, 129.4, 126.5, 126.3, 123.1, 123.0, 85.0, 69.1, 68.6, 68.3, 68.1, 67.5, 54.7, 41.2, 34.0, 32.3, 29.5, 29.46, 29.42, 29.0, 28.3, 27.8, 26.9, 24.6, 24.4. HRMS (TOF MS ES⁺) calcd for C₅₇H₆₄FeN₂O₄S₂Na 983.3556; found 983.3584.

N-(1,21-Bis(methyldisulfanyl)heneicos-11-yl)-*N*'-4-[1pyrenylmethyl])perylene-3,4,9,10-bis(dicarboximide) (1g). 1-NH₂ (21.97 mg, 0.095 mmol) and 7g (40.00 mg, 0.047 mmol) gave 1g (41 mg) in 82% yield as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.65-8.55 (m, 5H), 8.47-8.40 (m, 4H), 8.16-7.85 (m 8H), 6.09 (s, 2H), 5.20 (m, 1H), 2.65 (t, J = 6.0 Hz, 4H), 2.38 (s, 6H), 2.36-2.25 (m, 2H), 1.89 (m, 2H), 1.67-1.56 (m, 4H), 1.37-1.20 (m, 28H). ¹³C NMR (125 MHz, CDCl₃) δ 164.4, 163.2, 133.9, 133.5, 131.0, 130.8, 130.36, 130.33, 130.0, 128.9, 128.6, 128.5, 127.5, 126.9, 125.8, 125.58, 125.51, 125.4, 124.9, 124.8, 124.5, 124.3, 124.2, 122.9, 122.6, 122.3, 54.7, 41.2, 38.3, 32.4, 29.57, 29.51, 29.48, 29.42, 29.3, 29.1, 28.4, 27.0, 23.3. HRMS (TOF MS ES⁺) calcd for C₆₄H₆₆N₂O₄S₄Na 1077.3803; found 1077.3827.

N-(1,21-Bis(methyldisulfanyl)heneicos-11-yl)-*N*'-(2ferrocenylethyl)perylene-3,4,9,10-bis(dicarboximide) (3g). 3-NH₂ (21.76 mg, 0.095 mmol) and 7g (40.0 mg, 0.047 mmol) gave 3g (39 mg) in 79% yield as a waxy solid. ¹H NMR (300 MHz, CDCl₃) δ 8.64–8.46 (m, 8H), 5.24–5.19 (m, 1H), 4.40 (t, *J* = 9.0 Hz, 2H), 4.30–4.21 (m, 7H), 4.12 (m, 2H), 2.79 (t, *J* = 9.0 Hz, 2H), 2.67 (t, *J* = 6.0 Hz, 4H), 2.41 (s, 6H), 2.30–2.26 (m, 2H), 1.94–1.87 (m, 2H),

1.67–1.60 (m, 4H), 1.37–1.25 (m, 28H). ^{13}C NMR (75 MHz, CDCl₃) δ 164.3, 163.0, 134.4, 134.1, 131.7, 131.1, 130.9, 129.4, 129.1, 126.2, 126.1, 123.0, 122.98, 122.91, 85.1, 73.1, 69.6, 68.6, 68.1, 67.4, 54.8, 41.2, 38.3, 32.3, 29.5, 29.48, 29.42, 29.1, 28.4, 27.7, 26.9, 23.3. HRMS (TOF MS ES⁺) calcd for $C_{59}H_{68}\text{FeN}_2\text{O}_4\text{S}_4\text{Na}$ 1075.3311; found 1075.3282.

ASSOCIATED CONTENT

Supporting Information

¹H and ¹³C NMR spectra for new compounds. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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REFERENCES

(1) Wescott, L. D.; Mattern, D. L. J. Org. Chem. 2003, 68, 10058–10066.

- (2) Metzger, R. M.; Panetta, C. A. New J. Chem. 1991, 15, 209–221.
- (3) Mallouk, T. E.; Lee, H. J. Chem. Educ. 1990, 67, 829–834.
- (4) Aviram, A.; Ratner, M. A. Chem. Phys. Lett. 1974, 29, 277–283.
 (5) Metzger, R. M.; Mattern, D. L. Top. Curr. Chem. 2012, 313, 39–
- 84. (6) Shumate, J. W.; Mattern, D. L.; Jaiswal, A.; Dixon, D. A.; White,

 (d) Shumate, J. W., Wattern, D. L., Jaswai, K., Dixon, D. K., Winte,
 T. R.; Burgess, J.; Honciuc, A.; Metzger, R. M. J. Phys. Chem. B 2006, 110, 11146–11159.

(7) Martin, A. S.; Sambles, J. R.; Ashwell, G. J. Phys. Rev. Lett. 1993, 70, 218.

(8) Fan, F. R. F.; Yang, J. P.; Cai, L. T.; Price, D. W.; Dirk, S. M.; Kosynkin, D. V.; Yao, Y. X.; Rawlett, A. M.; Tour, J. M.; Bard, A. J. J. Am. Chem. Soc. **2002**, 124, 5550–5560.

(9) Majumder, C.; Mizuseki, H.; Kawazoe, Y. J. Phys. Chem. A 2001, 105, 9454.

(10) Mizuseki, H.; Niimura, K.; Majimder, C.; Kawazoe, Y. *Comput. Mater. Sci.* **2003**, *27*, 161–165.

(11) Metzger, R. M.; Chen, B.; Hopfner, U.; Lakshimikantham, M. V.; Ashwell, G. J. J. Am. Chem. Soc. **1997**, *119*, 10455.

(12) Ho, G.; Heath, J. R.; Kondratenko, M.; Perepichka, D. F.; Arseneault, K.; Pezolet, M.; Bryce, M. R. *Chem.—Eur. J.* **2006**, *10*, 2914–2922.

(13) Nazdizadeh, H.; Mattern, D. L.; Singleton, J.; Wu, X.; Metzger, R. M. Chem. Mater. **1994**, *6*, 268–277.

- (14) Halik, M.; Hirsch, A. Adv. Mater. 2011, 23, 2689-2695.
- (15) Cerruti, M.; Fissolo, S.; Carraro, C.; Ricciardi, C.; Majumdar, A.; Maboudian, R. *Langmuir* **2008**, *24*, 10646–10653.
- (16) Ashwell, G. J.; Chwialkowska, A.; High, H. J. Mater. Chem. 2004, 14, 2389–2394.

(17) Ashwell, G.; Mohib, A. J. Am. Chem. Soc. 2005, 127, 16238–16244.

(18) Ashwell, G. J.; Tyrrell, W. D.; Whittam, A. J. J. Am. Chem. Soc. 2004, 126, 7102–7110.

(19) (a) Demmig, S.; Langhals, H. Chem. Ber. 1988, 121, 225–230.
(b) Langhals, H.; Demmig, S.; Potrawa, T. J. Prakt. Chem. 1991, 333, 733–748.

(20) (a) Zhang, S.; Rio, Y.; Cardinali, F.; Bourgogne, C.; Gallani, J.-L.; Nierengarten, J.-F. J. Org. Chem. **2003**, 68, 9787–9797. (b) Iyer, J.; Hammond, P. T. Langmuir **1999**, 15, 1299–1306.

(21) Samudrala, R.; Žhang, X.; Wadkins, R. M.; Mattern, D. L. Bioorg. Med. Chem. 2007, 15, 186–193.

- (22) Wicklein, A.; Lang, A.; Muth, M.; Thelakkat, M. J. Am. Chem. Soc. 2009, 131, 14442–14453.
- (23) Cormier, R. A.; Gregg, B. A. Chem. Mater. 1998, 10, 1309–1319.
- (24) Williams, M. E.; Murray, R. W. Chem. Mater. 1998, 10, 3603–3610.
- (25) (a) Valkenier, H.; Huisman, E. H.; van Hal, P. A.; de Leeuw, D. M.; Chiechi, R. C.; Hummelen, J. C. J. Am. Chem. Soc. 2011, 133, 4930–4939. (b) Wang, C.; Batsanov, A. S.; Bryce, M. R. Faraday Discuss. 2006, 131, 221–234.

(26) (a) Jaiswal, A.; Rajagopal, D.; Lakshmikantham, M. V.; Cava, P.; Metzger, R. M. Phys. Chem. Chem. Phys. 2007, 9, 4007–4017.
(b) Girlando, A.; Sissa, C.; Terenziani, F.; Painelli, A.; Chwialkowska., A.; Ashwell, G. J. Chem. Phys. Chem. 2007, 8, 2195–2201.
(c) Béthencourt, M. I.; Srisombat, L.; Chinwangso, P.; Lee, T. R.

- Langmuir 2009, 25, 1265–1271. (27) Chen, L. X.; Xiao, S.; Yu, L. J. Phys. Chem. B 2006, 110, 11730– 11738.
- (28) Langhals, H.; Krotz, O.; Polborn, K.; Mayer, P. Angew. Chem., Int. Ed. 2005, 44, 2427–2428.
- (29) Kaiser, H.; Lindner, J.; Langhals, H. Chem. Ber. 1991, 124, 529-535.
- (30) Baker, R.; Herbert, R. H. J. Chem. Soc., Perkin Trans. 1 1987, 1123–1127.
- (31) Conway, J. C.; Quayle, P.; Regan, A. C.; Urch, C. J. *Tetrahedron* **2005**, *61*, 11910–11923.
- (32) Mudryk, B.; Cohen, T. *J. Am. Chem. Soc.* **1991**, *113*, 1866–1867. (33) Buffet, M. F.; Dixon, D. J.; Ley, S. V.; Reynolds, D. J.; Storer, R.
- I. Org. Biomol. Chem. 2004, 2, 1145-1154.
- (34) Arterburn, J. B.; Perry, M. C. Org. Lett. 1999, 1, 769-771.
- (35) Doubsky, J.; Streinz, L.; Saman, D.; Zednik, J.; Koutek, B. Org. Lett. 2004, 6, 4909–4911.
- (36) Reddy, G. B.; Mitra, R. B. Synth. Commun. 1986, 16, 1723–1729.
- (37) Kim, S.; Kavali, R. Tetrahedron Lett. 2002, 43, 7189-7191.

(38) Hamasaki, R.; Funakoshi, S.; Misaki, T.; Tanabe, Y. *Tetrahedron* 2000, *56*, 7423–7425.

- (39) Hironobu, M.; Sanda, F.; Endo, T. Macromolecules 2003, 36, 2206-2214.
- (40) Langhals, H. S.; Brandherm, M. T. Liebigs Ann. 1995, 481-486.
- (41) Tröster, H. Dyes Pigments 1983, 4, 171-177.
- (42) Pasaogullari, N.; Icil, H.; Demuth, M. Dyes Pigments 2005, 69, 118-127.
- (43) Nagao, Y.; Misono, T. Bull. Chem. Soc. Jpn. 1981, 54, 1191–1194.
- (44) Lattimer, R. P. J. Anal. Appl. Pyrolysis 2000, 56, 61-78.
- (45) Motesharei, K.; Myles, D. C. J. Am. Chem. Soc. 1997, 119, 6674–6675.
- (46) Pengo, P.; Broxterman, Q. B.; Kaptein, B.; Pasquato, L.; Scrimin, P. *Langmuir* **2003**, *19*, 2521–2524.
- (47) Davis, B. G.; Ward, S. J.; Rendle, P. M. Chem. Commun. 2001, 189–190.
- (48) Wysozogrodzka, M.; Möws, K.; Kamlage, S.; Wodzińska, J.; Plietker, B.; Haag, R. *Eur. J. Org. Chem.* **2008**, 53–63.
- (49) de Greef, T. F.; Nieuwenhuizen, M. M.; Sijbesma, R. P.; Meijer, E. W. J. Org. Chem. **2010**, 75, 598–610.
- (50) Brunner., H.; Schellerer, K. M. Inorg. Chim. Acta 2003, 350, 39–48.

(51) Chen, S.; Song, B.; Wang, Z.; Zhang, Xi. J. Phys. Chem. C 2008, 112, 3308–3313.

(52) Ravi, S.; Padmanabhan, D.; Mamdapur, V. R J. Indian Inst. Sci. 2001, 18, 299–312.

NOTE ADDED AFTER ASAP PUBLICATION

After ASAP publication on October 22, 2012, corrections were made to the experimental data for compounds **10**, **6b**, and **3b** in the Experimental Section; the correct version reposted October 23, 2012.